

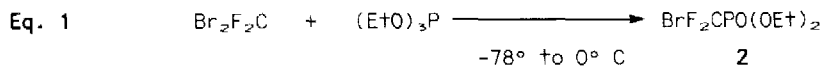
SYNTHESIS AND NMDA RECEPTOR BINDING OF
2-AMINO-7,7-DIFLUORO-7-PHOSPHONOHEPTANOIC ACID

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Abstract: In an NMDA specific receptor binding assay, **1** had lower affinity than its parent, APH. This result suggests that for competitive antagonists, diionization of the phosphonic acid moiety may be detrimental to receptor affinity.

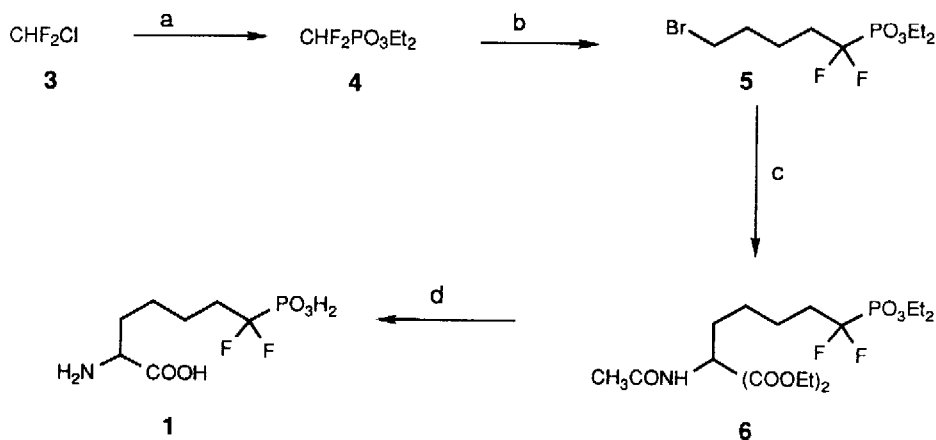
It is known that for competitive N-methyl-D-aspartic acid (NMDA) antagonists of the same overall structure a distal phosphonic acid moiety imparts a roughly ten fold increase in receptor binding affinity over that of the corresponding carboxylic acid.¹ Given the current rudimentary state of published NMDA receptor molecular modeling, the reason for this increase in affinity is unknown, but has been ascribed to the unique ability of phosphonic acid based antagonists to be diionized at physiological pH.² However, for a simple alkylphosphonic acid (pKa x and y) the second ionization will not be complete at physiological pH.³ This raises the possibility that additional features of the phosphonate group, other than diionization, may be responsible for the increased NMDA receptor binding affinity of known antagonists. To explore whether complete diionization is a prerequisite for increased receptor binding, we synthesized 2-amino-7,7-difluoro-7-phosphonoheptanoic acid **1**. In this derivative, the presence of the two proximal fluorine atoms was expected to lower the second pKa close to 6, thereby ensuring complete diionization at physiological pH,⁴ without significantly increasing the steric bulk α to the phosphonic acid.

Our initial attempts to prepare **1** centered on the conversion of dimethyl 1-oxo-5-chloropentylphosphonate into the corresponding α -difluoro derivative. Although precedented for α -ketoesters,⁵ treatment of the corresponding α -ketophosphonates with diethylaminosulfur trifluoride (DAST) under a range of conditions did not afford identifiable fluorinated products.⁶ As an alternative strategy in which both α -fluorines are already incorporated into a phosphonate synthon, we prepared diethyl (bromodifluoromethyl)phosphonate **2** via an Arbuzov reaction,⁷ Eq. 1.^{8,9} Although reported to undergo Reformatsky reactions with chloroacyl derivatives,¹⁰ and more recently cadmium mediated coupling with allyl bromide,¹¹ in our hands, **2** proved to be a poorly reactive substrate. Indeed, we were unable to generate and couple the zinc salt even with a simple activated halide such as benzyl bromide.



Diethyl (difluoromethyl)phosphonate **4** provided a more versatile synthon. Treatment of chlorodifluoromethane with sodium diethylphosphonate (Scheme 1) provided **4** as a volatile oil in good yield.¹²⁻¹⁴ Deprotonation with lithium diisopropylamide followed by addition to excess 1,4-dibromobutane at -78°C gave a moderate yield of the desired monoalkylated product **5**. Displacement of the remaining bromine with sodium acetamidomalonate proceeded smoothly to give **6**. Refluxing this product with 6 M hydrochloric acid resulted in the removal of all protecting groups and malonate decarboxylation to afford **1**.

SCHEME 1.



a) NaPO_3Et_2 , THF; b) LDA, THF, $\text{Br}(\text{CH}_2)_4\text{Br}$; c) $\text{NaC}(\text{NHAc})(\text{COOEt})_2$, EtOH, reflux; d) 6 M HCl, reflux

The pK_a values of the parent compound, 2-amino-7-phosphonoheptanoic acid (APH) and **1** were determined by potentiometric titration in water by first acidifying with HCl (aq), and then titrating with 0.2 N NaOH (aq). The observed dissociation constants for APH were $\text{pK}_{a_1} = 2.73$ and $\text{pK}_{a_2} = 7.89$, and for **1** were, as predicted, $\text{pK}_{a_1} = 3.0$ and $\text{pK}_{a_2} = 5.7$.¹⁵ Significantly, the increased dissociation of the second phosphonate proton of **1** compared with that of APH was illustrated both by facile phosphono monoester hydrolysis and the loss of HCl upon evaporation. Further, the non-typical physical properties of **1** are noted in the use of water as a recrystallization solvent. This is unusual due to the extreme hygroscopic nature of the non-fluorinated antagonists, and high water solubility of APH itself. These observations suggest that in **1** an internal salt exists between the phosphonic acid and the α -amine. Such an association might be expected to cause the molecule to fold in a manner which presents a predominately hydrophobic exterior to solvent, effectively screening its high intrinsic polarity.

Using [^3H]4-(3-phosphonopropyl)-2-piperazinecarboxylic acid (CPP) as a ligand, NMDA receptor binding was carried out as previously described.¹⁶ In this assay, APH had an IC_{50} of $0.8 \mu\text{M}$. Surprisingly, **1** had significantly less affinity for the NMDA receptor with an

IC_{50} of 27 μ M. The weak receptor binding affinity observed for **1** suggests that diionization of the distal phosphonic acid functionality may be detrimental to NMDA receptor binding. However, other factors may be responsible for the loss of receptor affinity. Although fluorine atoms have been used widely as replacements of hydrogen to modify activity in a variety of biologically active compounds,^{17,18} a change in the preferred conformation of **1** and/or adverse electronic interactions of the fluorine atoms with a receptor component may instead be responsible for this observed loss in receptor binding affinity. The existence of such an interaction cannot as yet, be determined.

Diethyl (5-bromo-1,1-difluoropentyl)phosphonic acid (5). A solution of **4** (5.6 gm, 0.03 mol) in THF (30 mL) was cooled in a dry ice-acetone bath, and treated via syringe with a solution of lithium diisopropylamide (20 mL, 1.5 M in cyclohexane). After stirring 30 min, the anion solution was cannulated into a cold solution (-78°C) of 1,4-dibromobutane (19.4 gm, 0.09 mol) in THF (70 mL). The reaction mixture was allowed to warm to room temperature, and then concentrated, quenched with 0.5 N HCl, and extracted with EtOAc (200 mL). The organic layer was washed with 1 N HCl (4 x 10 mL), saturated NaHCO₃ (20 mL), and saturated NaCl (20 mL). The EtOAc solution was dried over MgSO₄, filtered and evaporated. The oily residue was chromatographed on silica gel (50% EtOAc in heptane as eluant) to give **5** as a yellow oil (4.5 gm). MS (EI) 325 (40%), 323 (43%). ¹H-NMR (CDCl₃) 4H, m, 4.35-4.20 ppm; 2H, t, 3.42; 6H, m, 2.21-1.70; 6H, t, 1.38. Anal. (C₉H₁₆BrF₂O₃P) C, H, N.

(Acetylamino)[5-(diethoxyphosphinyl)-5,5-difluoropentyl]propanedioic acid diethyl ester (6). Diethylacetamidomalonate (2.94 g, 13.5 mmol) was added to a solution of sodium ethoxide (13 mmol) in EtOH (10 mL). After 30 min, **5** (3.5 g, 10.8 mmol) was added neat and washed into the reaction mixture with absolute EtOH (3 mL). The reaction mixture was refluxed overnight. The solvent was removed by evaporation and the residue was dissolved in EtOAc and washed consecutively with water and saturated NaCl solution. The organic layer was dried over MgSO₄, filtered and evaporated. The residue was chromatographed on silica gel (25-75% gradient of EtOAc in heptane as eluant) to give a white solid (1.8 g). The phosphonate monoethyl ester was obtained from the acidified aqueous wash as a light brown oil (1.5 g). MS (EI) 460 (82%). ¹H-NMR (CDCl₃) 1H, s, 6.78 ppm; 8H, m, 4.33-4.19; 2H, m, 1.72-1.55; 6H, t, 1.37; 6H, t, 1.25; 2H, m, 1.51-1.14. Anal. (C₁₈H₃₂F₂NO₈P) C, H, N.

(±)-2-Amino-7,7-difluoro-7-phosphonoheptanoic acid (1). **6** (1.5 g, 3.26 mmol) was refluxed in 6 N HCl (30 mL) overnight. The water was removed by evaporation in vacuo, and the residue was redissolved in water and lyophilized. The solid was washed with diethyl ether and recrystallized from water to give **1** as a white solid, (0.47 g). mp 240-241°C dec. MS (FAB) 262 (100%). ¹H-NMR (D₂O) 1H, t, 4.07 ppm; 4H, m, 2.18-1.92; 4H, m, 1.71-1.47. Anal. (C₇H₁₄F₂NO₅P) C calc'd 32.19; found 31.65, H, N.

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References

1. J. C. Watkins and H. J. Olverman in "Excitatory Amino Acids in Health and Disease" ed. D. Lodge, John Wiley and Sons, Chichester, UK., 1988, p. 13.
2. G. E. Fagg and J. Baud in "Excitatory Amino Acids in Health and Disease" ed. D. Lodge, J. Wiley and Sons, Chichester, UK., 1988, p. 63.
3. G. Kortum, W. Vogel and K. Andrussow, eds. "Dissociation Constants of Organic Acids in Aqueous Solution", International Union of Pure and Applied Chemistry, Butterworths, London, 1961. As an example of the dissociation constants of alkyl phosphonic acids, ethyl phosphonic acid has $pK_{a1} = 2.4$ and $pK_{a2} = 8.1$.
4. G. M. Blackburn and D. E. Kent, J. Chem. Soc. Perkin Trans. 1 1986, 913.
5. W. J. Middleton and E. M. Bingham, J. Org. Chem. 1980, 45, 2883; D. A. Trainor and M. M. Stein, EP 204571 A2, 1986.
6. Although monofluorination of an aromatic hydroxymethylphosphonate with DAST has been reported,⁴ the corresponding alkyhydroxymethylphosphonate afforded instead elimination or rearranged fluorinated products.
7. The Arbuzov reaction between dibromodifluoromethane and triethylphosphite should be treated with extreme caution. A violent reaction occurs (ca -20°C) which blew out the addition funnel, nitrogen inlet and stopper. This 'explosion' could be controlled by running the reaction in a huge flask, or more appropriately in a steel bomb.⁹
8. D. J. Burton and R. M. Flynn, J. Fluorine Chem. 1977, 10, 329.
9. T. Mahmood and J. M. Shreeve, Synthetic Commun. 1987, 17, 71.
10. D. J. Burton and L. G. Sprague, J. Org. Chem. 1988, 53, 1523.
11. R. D. Chambers, R. Jaouhari and D. Ohagan, J. Chem. Soc. Chem. Commun. 1988, 17, 1169.
12. L. Z. Soborovski and N. Baina, J. Gen. Chem. USSR 1959, 2, 1115.
13. M. Obayashi, E. Ito, K. Matsui and K. Kondo, Tetrahedron Letters 1982, 23, 2323.
14. C. E. McKenna and P. Shen, J. Org. Chem. 1981, 46, 4574.
15. The pK_{a1} and pK_{a2} values are apparently the dissociation constants for the α -carboxylic acid moiety and the second phosphonate proton. We were not able to detect a dissociation constant for the first phosphonate proton under the conditions used in this assay.
16. C. F. Bigge, J. T. Drummond, G. Johnson, T. Malone, A. W. Probert Jr., F. W. Marcoux, L. L. Coughenour and L. J. Brahce, J. Med. Chem. 1989, 32, 1580.
17. R. Filler, ed., "Biochemistry Involving Carbon-Fluorine Bonds", American Chemical Society, Washington D. C. 1978, ACS Symposium Series No. 28.
18. R. Filler and Y. Kobayashi, eds., "Biomedical Aspects of Fluorine Chemistry", Kodasha/Elsevier, New York, New York, 1982.

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